

NITROGEN FIXATION AND GROWTH RESPONSE OF *Alnus Rubra* FOLLOWING FERTILIZATION WITH UREA OR BIOSOLIDS

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ABSTRACT: Nitrogen fertilization of forests using biosolids offers a potentially environmentally friendly means to accelerate tree growth. This field study was designed to analyze the effects of nitrogen fertilization on the symbiotic, nitrogen (N)-fixing relationship between *Alnus rubra* Bong. (red alder) and *Frankia*. Anaerobically digested, class B biosolids and synthetic urea (46% N) were applied at rates of 140, 280 and 560 kg ha⁻¹ available N to a well-drained, sandy, glacial outwash soil in the Indianola series (mixed, mesic Dystric Xeropsamments). Plots were planted with *A. rubra* seedlings. At the end of each of two growing seasons trees were harvested and analyzed for the rate of N fixation (as acetylene reduction activity), biomass and foliar N. At year 1, there was no N fixation for trees grown with urea amendments, but control (17 $\mu\text{mol C}_2\text{H}_4 \text{ g}^{-1} \text{ hr}^{-1}$) and biosolids (26-45 $\mu\text{mol C}_2\text{H}_4 \text{ g}^{-1} \text{ hr}^{-1}$) trees were fixing N. At the end of year 2, all trees in all treatments were fixing N (7 $\mu\text{mol C}_2\text{H}_4 \text{ g}^{-1} \text{ hr}^{-1}$, 4-16 $\mu\text{mol C}_2\text{H}_4 \text{ g}^{-1} \text{ hr}^{-1}$, and 20-29 $\mu\text{mol C}_2\text{H}_4 \text{ g}^{-1} \text{ hr}^{-1}$ for control, urea and biosolids respectively). Trees grown with biosolids amendments were larger overall (year 1 shoot biomass 10 g, 5 g, and 23 g for control, urea, and biosolids respectively, year 2 shoot biomass 50 g, 51 g, and 190 g for control, urea, and biosolids respectively) with higher concentrations of foliar N for both years of the study (year 1 foliar N 26 g kg⁻¹, 27 g kg⁻¹, and 40 g kg⁻¹ for control, urea, and biosolids respectively, year 2 foliar N 17 g kg⁻¹, 19 g kg⁻¹, and 23 g kg⁻¹ for control, urea, and biosolids respectively). Trees grown with urea amendments appeared to use the urea N over *Frankia* supplied N, whereas the biosolids trees appeared to be able to use both N in biosolids and N from *Frankia*. The results from this study indicated that the greater growth of *A. rubra* may have been responsible for the observed higher N demand. Biosolids may have supplied other nutrients to the trees to support this accelerated growth.

Key words: *Frankia*, acetylene reduction activity (ARA)

FIXAÇÃO DE NITROGÊNIO E CRESCIMENTO DE *Alnus Rubra* FERTILIZADO COM URÉIA OU BIOSÓLIDOS

RESUMO: A fertilização nitrogenada de florestas com biosólidos constitui um meio de aceleração do crescimento das plantas potencialmente não impactante ao meio. Os efeitos de fertilização de nitrogênio atmosférico na relação simbiótica e de fixação de nitrogênio *Alnus rubra* Bong. (amieiro vermelho) e *Frankia* foram avaliados em um estudo de campo. Biosólidos classe B digeridos anaerobicamente e uréia sintética (46% N) foram aplicados a taxas de 140, 280 e 560 kg ha⁻¹ de N disponível em um solo tipo Areia Quartzosa. Ao contrário das plantas do grupo controle (17 $\mu\text{mol C}_2\text{H}_4 \text{ g}^{-1} \text{ hr}^{-1}$) ou fertilizadas com biosólidos (26-45 $\mu\text{mol C}_2\text{H}_4 \text{ g}^{-1} \text{ hr}^{-1}$), plantas fertilizadas com uréia não apresentavam fixação de nitrogênio após um ano de cultivo. Ao final do segundo ano, todas as árvores em todos os tratamentos fixavam N (7 $\mu\text{mol C}_2\text{H}_4 \text{ g}^{-1} \text{ hr}^{-1}$; 4-16 $\mu\text{mol C}_2\text{H}_4 \text{ g}^{-1} \text{ hr}^{-1}$; e 20-29 $\mu\text{mol C}_2\text{H}_4 \text{ g}^{-1} \text{ hr}^{-1}$ para controle, uréia e biosólidos, respectivamente). Plantas cultivadas sob remediação com biosólidos apresentaram maior biomassa na parte aérea ao final do ano 2 (50 g; 51 g; e 190 g para controle, uréia, e biosólidos, respectivamente), e também maiores concentrações de nitrogênio foliar em ambos os períodos analisados (N foliar no ano 1 – 26 g kg⁻¹, 27 g kg⁻¹, e 40 g kg⁻¹; N foliar no ano 2 – 17 g kg⁻¹, 19 g kg⁻¹, and 23 g kg⁻¹, respectivamente para controle, uréia e biosólidos). A maior taxa de crescimento de *A. rubra* pode ter resultado em maior exigência em N e os biosólidos supriram quantidades adicionais de nutrientes capazes de sustentar o crescimento acelerado.

Palavras-chave: *Frankia*, atividade da redução de acetileno (ARA)

INTRODUCTION

Traditionally, commercial forestry has been managed without consideration of potential impacts on forest ecosystems. One example of this is the impact of forest nitrogen (N) fertilization, with synthetic fertilizers or municipal biosolids, on the atmospheric N fixation in forests. *Alnus rubra* is an important hardwood in forests on the Pacific Coast of North America and is frequently found in *Pseudotsuga menziesii* (Douglas fir) plantations. *A. rubra* forms a symbiotic N fixing relationship with *Frankia* which complicates the determination of appropriate rates of synthetic N or biosolids N to maximize growth while simultaneously preventing environmental contamination (Henry et al., 2000).

It is difficult to predict how *A. rubra* will react to the addition of N from biosolids, as there has been scant research on fertilization of *Alnus* sp. with N. For other plant species that form symbiotic N fixing relationships, the addition of N fertilizer has been shown to slow or stop N fixation rates (Voison et al., 2002), which then increase upon the subsequent removal of the N source (Fujikake et al., 2002). Available N can also reduce nodule frequency and size (Daimon & Yoshioka, 2001). There have been studies illustrating similar effects with *Alnus* (Côté & Dawson, 1989; Martin et al., 2003). However, there is also the potential that *Alnus* would utilize both symbiotic and added sources of N if plant demand for N was sufficiently strong (Troelstra et al., 1992). Biosolids soil amendments have increased growth and nodulation of *Vicia sativa* (Vetch) (Sidiras et al., 1999), and *Glycine max* (soybeans) (Vieira, 2001). In studies looking at *Alnus*, N additions as NO_3^- , NH_4^+ , and NH_4NO_3 increased biomass of *A. glutinosa* (Troelstra et al., 1992), and N additions did not reduce N fixation in *A. incana* unless the system N demand was exceeded (Rytter et al., 1991).

This study was conducted to evaluate what the effect of N from urea and biosolids would be on the symbiotic N fixing relationship of *Alnus-Frankia*, and if increased growth yields would result from these N additions.

MATERIALS AND METHODS

Study Site Description

This study was conducted at a research forest in Eatonville, Washington, US (47°37'N, 122°19'W) at the base of the Cascade Mountain foothills. Average annual rainfall is 1200 mm, with the rainfall from July and August typically being less than 120 mm (Pack Forest precipitation records). The study site is 317 m

above sea level. The soil is a well-drained, sandy, glacial outwash soil in the Indianola series (mixed, mesic Dystric Xeropsamments) (NRCS, 2004).

Study Installation

Prior to installation in February 2002 the site was densely vegetated with *Cytisus scoparius* (Scotch Broom) and *A. rubra*. The area was cleared of vegetation and the soil was disked to break up any remaining roots for removal. Nitrogen was provided in two forms; synthetic urea [46% N ($\text{CO}(\text{NH}_2)_2$)] (urea), and class B, anaerobically digested biosolids with available and total N content of 14 g NH_3 kg^{-1} and 63 g N kg^{-1} , respectively (King County Environmental Laboratory, March 2002). A control treatment was also included in the experimental design.

Biosolids application rates were calculated based on the method used by King County for their biosolids forest application sites (Henry et al., 1999). This method considers N uptake by trees and understory, soil immobilization and potential for volatilization. For this site consideration of these factors resulted in a biosolids application of 13.4 Mg ha^{-1} which provided 140 kg ha^{-1} available N. In comparison, recommended available N rates for the first two years growth of hybrid cottonwood (*Populus* L.; do not fix N) are 195 kg ha^{-1} (Henry et al., 1999). In addition to the rate calculated to meet the N needs of the stand, biosolids and urea were added at 2x and 4x this amount. The study was designed as a randomized complete block with three replicates. Plots were each 2 m \times 2 m. There was a 1 m space between adjoining plots as well as a 3 m work row between each block. The closest distance between trees in adjoining plots was 2 m.

Amendments were applied evenly to the surface of each plot and then rototilled to a depth of 15–20 cm. Nine bare-root *A. rubra* Bong. seedlings, 40–50 cm in height, were planted in each plot evenly spaced in a square grid pattern, with a 0.5 m buffer between the outermost trees and the edge of the plot. There were 27 trees planted for each treatment (9 per plot \times 3 plots per treatment). The seedlings were obtained from a local nursery and had been grown from seed stock from lowlands in Whatcom and Skagit counties. At the time of planting, the whips had sparse and small nodules and no leaves.

Maintenance

To eliminate any additional competition for soil available N weeds within each plot area were killed by direct application of glyphosate herbicide to foliage with a wick applicator. The area surrounding plots was mowed to discourage additional weeds. Deer browsing of *A. rubra* foliage was initially controlled by bio-control methods. In December 2002, a 2.5 m chain

link fence was built surrounding the entire study and this eliminated access for deer. After the first growing season, plots were thinned so that a maximum of 4 trees remained in each plot. This was done to reduce competition between trees as they grew.

Soil/Biosolids Sampling

Soil samples were collected prior to amendment application, as well as during tree harvest in September 2002 and August 2003. Soil samples were collected with a stainless steel probe (2.5 cm diameter) in the 0–15 cm depth range. Three samples were taken from each plot and homogenized. Samples were kept on ice or refrigerated until return to the laboratory when the available N analysis was performed using field moist samples that were sieved to <2 mm; all other analyses were conducted using air-dried samples that had been sieved to <2 mm. Prior to installation total carbon (C) and N were determined by combustion and total phosphorus (P) was determined by soil acid–peroxide digestion method for sediments, sludges, and soils (USEPA, 1986) on a per plot basis. Average C content of soils prior to amendment addition was $2 \pm 0.3 \text{ g kg}^{-1}$. Total N content was $0.1 \pm 0.001 \text{ g kg}^{-1}$ with total P equal to $428 \pm 25 \text{ g kg}^{-1}$.

Available Nitrogen and phosphorus

Potassium chloride (KCl) extraction was used to determine available N (NO_2^- , NO_3^- and NH_4^+) (Keeney & Nelson, 1982; Robertson et al., 1999). Five grams of soil were placed in a glass flask with 50 mL 2 M KCl. The flasks were placed on a shaker for one hour, left to settle, and then filtered through Whatman No. 41 filter paper. The solution was analyzed with a Lachat flow injection analyzer using QuickChem method 107-04-1-B for $\text{NO}_3^-/\text{NO}_2^-$, and 10-107-06-1-F for NH_4^+ . Ammonium nitrate (NH_4NO_3) solution standards were used to calibrate for solution concentrations of NO_3^- and NH_4^+ from 0 to $1000 \mu\text{g L}^{-1}$ (calibration curve coefficient = 0.99). Soil water content data were used to correct for percent moisture of field moist samples.

Available P was determined using the Bray method (Kuo, 1996). Twenty mLs of dilute acid solution were added to 1 g of air dried soil. The mixture was shaken for 2 minutes and then filtered through Whatman No. 40 filter paper. Total P in the extracts was determined on a Lachat flow injection analyzer.

pH

Soils slurries were mixed 1:1 on a volume basis with deionized water and air-dried soil samples and then left to stabilize. The pH was measured after one hour with an Orion pH meter (Bremner, 1996; Nelson & Sommers, 1996).

Total Carbon and Nitrogen

Soil samples were ground with a mortar and pestle and placed into foil packages on a CAHN C-33 microbalance. Total C and N were determined using a CE-440 elemental analyzer, at 980°C (Bremner, 1996; Nelson & Sommers, 1996). National Institute of Standards and Technology (NIST) standards “Montana soil” and “sludge standard”, along with acetanilide and benzoic acid as internal standards, were routinely included in analysis. Respective rates of recovery for standards were; 0.89–0.96 (C, Montana soil), 0.95–1.06 (N, sludge standard), 0.94–1.01 (C, acetanilide), 0.94–0.96 (N, acetanilide), 0.98 (C, benzoic acid). Total C and N were reported as % based on a dry weight basis, and the C:N ratio as weight:weight.

Nitrogen Fixation

There are advantages and deficiencies to every method for measurement of symbiotic N fixation. Methods reported in the literature include ^{15}N isotope dilution (Busse, 2000), ^{15}N natural abundance (Shearer et al., 1983), ^{15}N incorporation (McNeill et al., 1997), N accretion (Rothe et al., 2002), and acetylene reduction activity (ARA) (Hardy et al., 1973). As the goal of this study was to evaluate the impact of treatments on *Frankia*, and not to determine cumulative amounts of N fixation, ARA was selected as the most convenient method of analysis (Myrold et al., 1999). ARA has been shown to be a sensitive method for comparison of relative rates of N fixation (Weaver & Danso, 1994). Other methods such as isotope dilution are complicated by the need for a non-fixing reference plant that has similar growth patterns to the unique, rapid growth habits of *Alnus* spp. (Busse, 2000). *In situ* methods (e.g. split-root systems, cuvettes) are more readily carried out in the laboratory and are not necessarily appropriate for field studies. For this study, foliar N was used in addition to ARA to evaluate the nitrogen balance between control and fertilized treatments.

Relative rates of N fixation were measured as ARA and reported as micromoles C_2H_4 produced per gram of nodule used in the assay (dry weight basis) per hour ($\mu\text{mol C}_2\text{H}_4 \text{ g}^{-1} \text{ h}^{-1}$) as described in Hardy et al. (1973). Two trees from each plot were harvested for analyses during the first week of September in 2002, and the last week of August in 2003. The values from both trees were averaged and treated as a single sample for statistical analysis. This time of year was selected for tree harvesting to allow for a reasonably complete growing season for comparison of biomass. In addition, it was determined based on reported midday observations of *A. rubra* ARA in July ($34.9 \mu\text{mol g}^{-1} \text{ hr}^{-1}$), September ($20.9 \mu\text{mol g}^{-1} \text{ hr}^{-1}$) and Janu-

ary ($0.1 \mu\text{mol g}^{-1} \text{hr}^{-1}$) (Teklehaimanot & Martin, 1999) that ARA would be sufficiently high at this time. Replicate blocks were analyzed on consecutive days at the same time of the day to account for diurnal variations. Tree roots were dug up with a garden fork and soil was lightly brushed off. A random selection of nodules was removed from roots with a razor knife, keeping a portion of the root intact to ensure that damage did not occur to nodules, and placed into a 65 mL glass vial, which was then sealed with rubber septa. 6.5 mL air was removed from the vial and then 6.5 mL acetylene (C_2H_2) was added (10% by volume), and the head space was mixed well. Positive controls (nodules, no C_2H_2) and negative controls (C_2H_2 , no nodules) were included. The reaction was left to run for three hours in the dark and then a 3ml sample was taken from each vial and stored in a vacutainer for transport to the laboratory. Samples were analyzed for ethylene (C_2H_4) on a SRI 8610C gas chromatograph (GC) with a $60 \text{ m} \times 0.53$ Supelco capillary column, using manual injection with nitrogen as the carrier gas (Hardy et al., 1973; Myrold et al., 1999; Weaver & Danso, 1994). Detection limits for this analysis, taking dilution into account, were $1.5 \mu\text{mol g}^{-1} \text{hr}^{-1}$. Soil moisture and temperature were also determined for each plot at the time of ARA sampling to account for potential variability.

Biomass

Whole trees were harvested for biomass determination. Shoots and roots were separated at the soil line for root and shoot biomass measurements in 2002. All nodules were brushed to remove soil and removed from roots for separate biomass measurements. Although multiple plants were harvested per plot, the biomass measures of the plants were averaged and a single average value was used for statistical analysis. Plants were oven-dried for 72 hours at 70°C . Biomass was reported as dry weight in grams. Two trees were harvested from each plot the first year of the study. For the second year only shoots were harvested as dry cemented soil conditions made it difficult to accurately collect the entire root and nodule system for determination of total biomass. Mean \pm standard deviation is reported in the text.

Total Foliar Nitrogen

Three leaves were randomly selected from each tree to comprise one composite sample for both trees collected from each plot. Samples were oven-dried, ground and then analyzed for total N on a CE-440 elemental analyzer as described for soils (Bremner, 1996; Nelson & Sommers, 1996). NIST plant standards as well as acetanilide and benzoic acid as internal standards were included in analysis. Respective rates of recovery for standards were; 0.96-1.01 (N,

citrus leaves), 0.94-1.01 (C, acetanilide), 0.94-0.96 (N, acetanilide), 0.98 (C, benzoic acid). Mean \pm standard deviation is reported in the text.

Statistical Analysis

Statistical analyses were performed using SPSS 12 for Windows (SPSS Inc, 2003). Data was checked for normality by visual inspection and confirmed using Kolmogorov-Smirnov tests. Analysis of variance (ANOVA) was used to test for main effects of treatment and treatment versus rate interactions. There was a small block effect for both foliar N and total soil C the first year and no significant block effects for the second year of the study. There were no block versus treatment interactions for either year. Means were separated using Duncan's multiple range test ($P = 0.05$). Linear regression was used to evaluate relationships between treatment rate and measured properties, and correlation was evaluated among measured properties ($P = 0.05$).

RESULTS AND DISCUSSION

Soil Properties

At the end of the first growing season (Table 1) both urea and biosolids treatments had higher levels of available N than the control. NO_3^- concentrations were similar for biosolids and urea for each treatment, with soil NH_4^+ concentrations overall being lower for urea than biosolids. As expected, almost the entire N from urea application was in the form of plant available N (PAN) for the first year of the study and had dissipated by the second year. There was a slight decrease in the C:N ratio for almost every treatment from the first to the second year. The first year soil pH values for urea and biosolid treatments were both lower than that of the control, while in the second year the biosolids pH was lower than both the control and urea. The lowest pH was in the year 2 biosolids 560 kg ha^{-1} treatment (pH = 4.9) as compared to the highest in the year 2 control (pH=5.8). Plant available P was higher in all of the biosolids treatments than in the control or urea treatments. This is to be expected as biosolids contain high levels of P in addition to N. The extractable P in the biosolid amended soils are considered optimal for plant growth, whereas the P in the urea and control soils were below optimal levels, potentially indicating deficiency (SERA-IEG 17, 2000).

Nitrogen fixation and nodule biomass

At the time of the first year tree harvest, N fixation activity as ARA was below detection limits for all trees grown with urea amendments (Figure 1). Rates of ARA for trees grown in biosolid treatments ($26\text{-}45 \mu\text{mol C}_2\text{H}_4 \text{ g}^{-1} \text{hr}^{-1}$) were higher than control trees (17

Table 1 - Mean \pm standard deviation for soil NO_3^- , NH_4^+ , total C, N, and C:N by treatment type and rate for both years. Biosolids were applied at rates to provide equivalent available N as the Urea.

		Year 1						
Treatment		NO_3^-	NH_4^+	C	N	C:N	pH	P
mg kg^{-1} avail. N		mg kg^{-1}		g kg^{-1}				mg kg^{-1}
Control	0	3 ± 0	3 ± 2	0.20 ± 0.01	0.01 ± 0.00	5 ± 0	5.6 ± 0.3	12.8 ± 4.5
Urea	140	35 ± 28	5 ± 4	0.21 ± 0.01	0.05 ± 0.00	4 ± 0	5.4 ± 0.4	14.7 ± 5.9
Urea	280	50 ± 32	7 ± 5	0.21 ± 0.08	0.04 ± 0.01	6 ± 2	5.1 ± 0.2	12.4 ± 2.7
Urea	560	69 ± 24	6 ± 3	0.23 ± 0.09	0.05 ± 0.02	4 ± 1	4.9 ± 0.0	13.1 ± 1.9
Biosolids	140	39 ± 18	19 ± 19	0.33 ± 0.08	0.07 ± 0.03	5 ± 1	5.1 ± 0.1	23.1 ± 2.3
Biosolids	280	39 ± 7	17 ± 12	0.26 ± 0.11	0.05 ± 0.01	6 ± 2	5.2 ± 0.0	25.2 ± 3.1
Biosolids	560	70 ± 27	65 ± 28	0.27 ± 0.08	0.06 ± 0.02	4 ± 1	5.2 ± 0.1	51.4 ± 29

		Year 2						
Treatment		NO_3^-	NH_4^+	C	N	C:N	pH	
mg kg^{-1} avail. N		mg kg^{-1}		g kg^{-1}				
Control	0	0.4 ± 0.4	1.5 ± 1.5	0.20 ± 0.04	0.10 ± 0.01	2 ± 1	5.8 ± 0.1	
Urea	140	0.8 ± 0.8	2.7 ± 0.1	0.26 ± 0.14	0.08 ± 0.01	3 ± 2	5.5 ± 0.1	
Urea	280	0.1 ± 0.0	0.7 ± 0.2	0.20 ± 0.05	0.09 ± 0.01	2 ± 0	5.6 ± 0.1	
Urea	560	2.6 ± 1.3	6.0 ± 3.5	0.24 ± 0.14	0.09 ± 0.04	3 ± 2	5.3 ± 0.2	
Biosolids	140	0.9 ± 0.1	1.1 ± 0.6	0.30 ± 0.05	0.09 ± 0.01	4 ± 1	5.5 ± 0.0	
Biosolids	280	2.7 ± 1.8	3.4 ± 2.7	0.27 ± 0.06	0.10 ± 0.02	3 ± 0	5.3 ± 0.1	
Biosolids	560	19 ± 17	16 ± 12	0.32 ± 0.09	0.08 ± 0.03	4 ± 1	4.9 ± 0.2	

$\mu\text{mol C}_2\text{H}_4 \text{ g}^{-1} \text{ hr}^{-1}$), but high variability precluded these differences from being statistically defensible ($P = 0.05$). Nodule biomass followed a similar pattern to ARA with the trees grown in urea ($0.05 \pm 0.03 \text{ g}$) and control treatments ($0.13 \pm 0.06 \text{ g}$) being smaller, and biosolid treatments larger ($0.20 \pm 0.10 \text{ g}$). In the first year total nodule biomass correlated positively to ARA ($r^2 = 0.60$). However, there was no relationship between the amount of amendment to nodule biomass or ARA. The absence of any detectable N fixation (as ARA), together with the smaller nodule biomass of trees grown in urea amended soils, was most likely a result of available N in the soil inhibiting N fixation and nodulation. This effect has been well documented for *Alnus* species (Huss-Danell et al., 2001; Martin et al., 2003; Wall et al., 2000).

At the time of the second year tree harvest, when N had dissipated from the urea treatments, rates of N fixation in the urea treatments increased to 4-16 $\mu\text{mol C}_2\text{H}_4 \text{ g}^{-1} \text{ hr}^{-1}$. ARA for control trees was 7 $\mu\text{mol C}_2\text{H}_4 \text{ g}^{-1} \text{ hr}^{-1}$ and biosolids 20-29 $\mu\text{mol C}_2\text{H}_4 \text{ g}^{-1} \text{ hr}^{-1}$. Differences between type of treatments were not statistically different ($P = 0.05$), but ARA activity in the urea amendments increased proportionally with the amount of treatment applied in the first year ($r^2 = 0.70$, $P = 0.05$). Where ARA was inhibited the first year for the urea trees, the second year trees were fixing N at rates similar to the control soils. ARA for trees grown

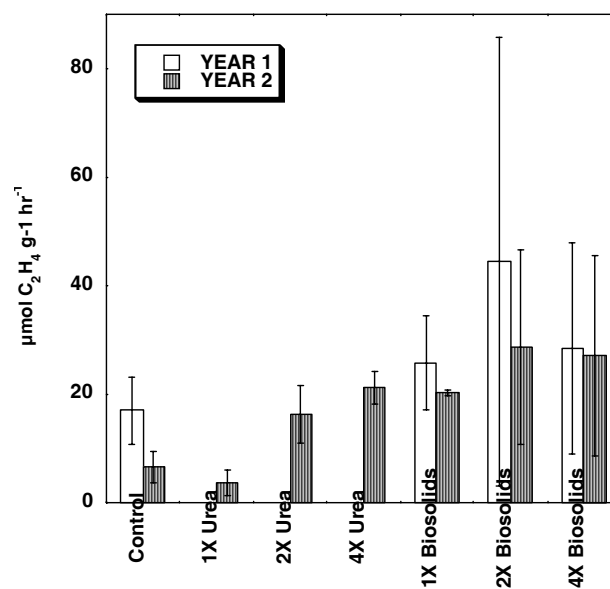


Figure 1 - Nitrogen fixation by *Frankia* as measured by acetylene reduction activity (ARA) for *Alnus rubra* fertilized with urea or municipal biosolids applied to provide one, two or four times the estimated N demand. Data, means and standard deviation are shown for both years.

in the biosolid treatments were not inhibited by available N in either year of the study. This was the case even though the biosolids and urea amendments were designed to provide the same amounts of available N. These results can potentially be explained by consid-

ering the other nutrients included in the biosolids. In particular, N, P and the ratio between them have been found to increase nodulation and N fixation in actinorhizal species. Studies growing actinorhizal species in solution have found N to be an inhibitor and P a promoter of nodule growth (Wall et al., 2000). In addition, at low P concentrations such as those observed in the control and urea treatments, soil N inhibits nodule size and number, but at high P concentrations N stimulates nodule growth (Gentili & Huss-Danell, 2003; Huss-Danell et al., 2001). In a field study on the Tanana River floodplain N fixation of *A. tenuifolia* was found to be P limited at total P concentrations of 79 g m⁻² (depth of 20 cm) (Uliassi & Ruess, 2002). Phosphorus fertilization increased nodule biomass and resulted in a 138% increase in *A. tenuifolia* N fixation. Based on the N fixation observed in the biosolid treatments, our results suggest that the P in the biosolids was a factor in increasing N fixation.

Shoot biomass

The average shoot biomass at the time of the first years sampling were 5 ± 2 g for trees grown in urea and 10 ± 8 g for the control trees, with the biosolids trees being larger 23 ± 11 g than the urea and control trees (Figure 2). There was a weak correlation between total biomass and rates of ARA ($r^2 = 0.49$). At the second year harvest the differences in shoot biomass between trees grown in the biosolids amended soils and the other treatments were again very pronounced, with the control and urea treatments be-

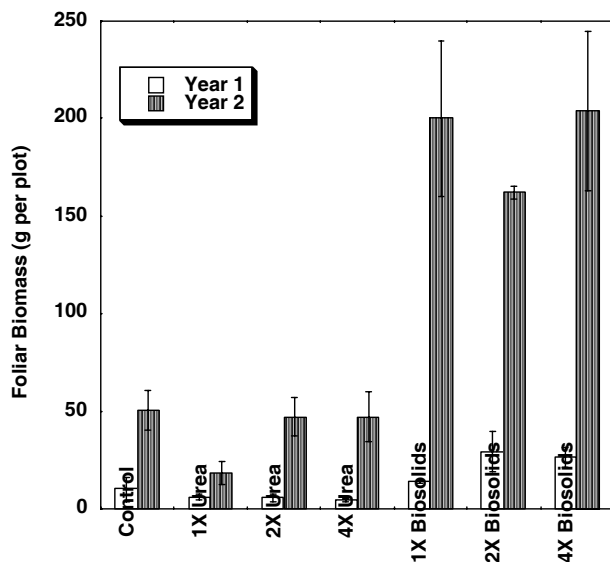


Figure 2 - Foliar biomass for *Alnus rubra* fertilized with urea or municipal biosolids applied to provide one, two or four times the estimated N demand. Means and standard deviation are shown for both harvests with two trees harvested per plot per year.

ing smaller (190 ± 45 g biosolids, 50 ± 14 g control and 51 ± 38 g urea). No correlation was seen between shoot biomass and ARA, although in other studies *A. rubra* biomass has frequently been seen to correlate to ARA (Monaco et al., 1982; Rojas et al., 2002).

Shoot biomass decreased with increasing amount of urea amendment, while shoot biomass of trees grown with biosolids amendments were 2 to 5 times larger than control and urea treatments. Observed yield increases were similar across all rates of biosolid additions. Total P in the biosolids used in this study was 26.7 g kg⁻¹ and plant available P in all biosolid treatments was greater than or equal to 25 mg kg⁻¹. Plant available P in all other treatments averaged 13.3 mg kg⁻¹ which is below optimal levels for plant growth (SERA-IEG 17, 2000). At the lowest application rate (6 Mg ha⁻¹), the biosolids provided 161 kg P ha⁻¹. Although recent studies have demonstrated that the phytoavailable fraction of total P in biosolids varies based on biosolids treatment, results from this study suggest that P, or other nutrients provided by the biosolids, may have been the factor responsible for the high growth and subsequent high N demand in trees grown in these treatments (O'Connor et al., 2004).

A growth response has been seen when P is added to nitrogen fixing plants, but it is unknown whether or not the P limitation is due to nodulation and nodule function of the symbiotic relationship (Gentili & Huss-Danell, 2003; Olivera et al., 2004), or the host plant demand for P (Reddell et al., 1997). Binkley et al. (2003) found P fertilization of *Facaltaria moluccana* (actinorhizal tree) to increase both the seedling biomass by 75% and N fixation by 100%. Rojas et al. (2002) found the addition of macronutrients (P, K, Ca, Mg), combined with inoculation of *A. rubra* with *Frankia*, to also increase both rates of ARA and biomass.

Total P concentration in the soil for this study was 428 mg kg⁻¹ and phytoavailable P was below optimal levels. In another study also conducted on a gravelly, sandy loam, Indianola series soil, Compton & Cole (1998) found total soil P in a 50 year-old *A. rubra* stand that colonized following a clear cut of *P. menziesii* / *Tsuga heterophylla* (Douglas fir/western hemlock) forest to be between 940 and 1130 mg kg⁻¹ in the upper 15 cm of soil.

The first year root:shoot ratio of biosolid amended trees (0.45 ± 0.07) and control trees (0.47 ± 0.09) were both lower than the urea treatment (0.60 ± 0.13). The average total tree biomass of the biosolid, control and urea treatment trees was; 33 ± 14 g, 14 ± 11 g and 7 ± 3 g, respectively. Although Troelstra et al. (1992) found the addition of available N to increase

the biomass of *A. incana*, during the first year of this study adding urea to the soil actually reduced tree growth, and it appears that at the highest rate of urea application the reduced yield seen in this study may have been partially related to NH_3 toxicity.

Foliar N

During both years of the study foliar N was higher in trees grown in the biosolids amended soils than other treatments (Figure 3). This difference was most pronounced during the first year of the study with foliar N in trees grown with biosolids averaging $40 \pm 1 \text{ g kg}^{-1}$ for comparison to $26 \pm 1 \text{ g kg}^{-1}$ for the control and $27 \pm 1 \text{ g kg}^{-1}$ for the urea treated soils. Foliar N for trees in urea treatments decreased with increasing rate of amendment. The first year trees growing in the 140 and 280 kg ha^{-1} urea treatments, which had no detectable ARA activity, also had equivalent foliar N concentrations in relation to control trees indicating that the trees were able to utilize added soil available N. However, decreased foliar N of trees grown in the 560 kg ha^{-1} urea treatment, along with reduced shoot biomass, may again indicate NH_3 toxicity.

For the second year, trees grown in biosolids amended soil had foliar N concentrations of $23 \pm 2 \text{ g kg}^{-1}$ as compared to $17 \pm 3 \text{ g kg}^{-1}$ for the control and $19 \pm 3 \text{ g kg}^{-1}$ for the urea amended soils. Foliar N concentration for trees grown with urea were similar to the control during the second year of the study when

available N was depleted from the soil. Second year foliar N, in combination with higher rates of ARA activity, indicate that at this time *Frankia* was supplying N for the urea trees. The first year foliar N results for all trees were higher than the second year when soil N was also higher, indicating that *Alnus* were utilizing added soil N. The higher foliar N, rates of N fixation and shoot biomass of trees grown with biosolid amendments indicate that the N demand of trees was not being met entirely by *Frankia* or biosolids, but by a combination of *Frankia* and biosolids N concurrently. Urea trees appeared to only utilize either urea or *Frankia* N, not a combination of the two, the reduced growth and resulting N demand of the trees in the urea treatments may have been the result of a nutrient deficiency. In comparison, Compton et al. (1997) did not see a difference in foliar N when comparing two stands of *A. rubra* between soils with differing P and pH, indicating that *Frankia* was able to supply the difference, but did find trees from lower P soils to also have lower foliar P, Ca, and Mg in addition to lower shoot biomass. In another study, Beaupied et al. (1990) found that the majority of leaf N in *A. glutinosa* came from symbiotically fixed N, even if there was available N in the soil, as shown by rates of N fixation and soil mineral N concentrations. Levels of available N in that study ($\sim 2\text{-}20 \text{ mg NO}_3^- \text{ kg}^{-1} \text{ soil}$, $\sim 2\text{-}12 \text{ mg NH}_4^+ \text{ kg}^{-1} \text{ soil}$) were lower than levels in this study.

CONCLUSION

The *Alnus/Frankia* relationship for responses to N added in the different forms of urea and biosolids showed positive effects resulting from the addition of biosolids and little response to added urea. Biosolids treated trees had higher shoot biomass, foliar N, and rates of N fixation than the urea and control trees. Urea inhibited rates of N fixation the first year, and had a negative impact on growth response. For low productivity soils, land application of biosolids to *A. rubra* appears to be an efficient method to maximize yield response. Although it was beyond the scope of this study to measure effects of NO_3^- leaching, the high growth rate observed by trees grown in the biosolids amended soils suggests that N demand may have been high enough to reduce the potential for nitrate leaching.

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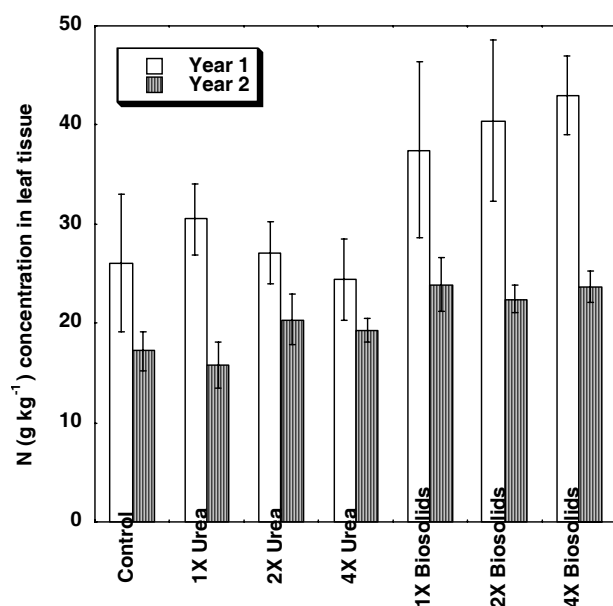


Figure 3 - Foliar N for *Alnus rubra* collected for both years of the study. Means \pm standard deviation are presented. Trees were fertilized with urea or municipal biosolids applied to provide one, two or four times the estimated N demand prior to planting.

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